

Treatment for Spinal Muscular Atrophy Using Onasemnogene Apeparovvec

Can Ebru Bekircan-Kurt,¹ Megan A Waldrop,^{1,2} Anne M Connolly¹ and Jerry R Mendell²

1. Center for Gene Therapy, Nationwide Children's Hospital, Columbus, OH, USA; 2. Department of Pediatrics, Nationwide Children's Hospital, Columbus, OH, USA

Spinal muscular atrophy (SMA) is the most common cause of death in infancy. Recently introduced molecular-based approaches have changed the poor prognosis, saved lives and improved the quality of life for those affected with SMA. Gene therapy uses an adeno-associated virus (AAV) to deliver and replace the mutant survival of motor neuron (*SMN*) genes, *SMN1* and *SMN2*. This review describes the development, relative safety and efficacy of intravenously delivered AAV for SMA type 1 and the intrathecal delivery for SMA type 2. For SMA, viral immunosuppressive treatment and AAV doses never used in clinical research or practice were required for success. As a prototype, the approach has greatly influenced the development of treatment for other childhood and adult diseases. Two additional pharmacologic agents, nusinersen and risdiplam, are clinically approved as alternative treatments. Both use antisense oligonucleotides and are briefly described in this review.

Keywords

Adeno-associated virus, gene therapy, nusinersen, onasemnogene abeparovvec, risdiplam, SMA type 1, spinal muscular atrophy

Disclosures: Jerry R Mendell, Anne M Connolly and Megan A Waldrop participate in research for SMA clinical trials supported by Avexis and Novartis Pharmaceuticals. Can Ebru Bekircan-Kurt has no financial or non-financial relationships or activities to declare in relation to this article.

Review process: Double-blind peer review.

Compliance with ethics: This article involves a review of the literature and did not involve any studies with human or animal subjects performed by any of the authors.

Data availability: Data sharing is not applicable to this article as no datasets were generated or analysed during the writing of this article.

Authorship: The named authors meet the International Committee of Medical Journal Editors (ICMJE) criteria for authorship of this manuscript, take responsibility for the integrity of the work as a whole, and have given final approval for the version to be published.

Access: This article is freely accessible at touchNEUROLOGY.com. © Touch Medical Media 2022

Received: 17 August 2022

Accepted: 23 September 2022

Published online: 23 November 2022

Citation: *touchREVIEWS in Neurology*. 2022;18(2):Online ahead of journal publication

Corresponding author: Jerry R Mendell, 700 Children's Drive, Nationwide Children's Hospital and The Ohio State University, Columbus, OH 43205, USA. E: Jerry.Mendell@nationwidechildrens.org

Support: No funding was received in the publication of this article.

Background for molecular-based therapy for spinal muscular atrophy

Spinal muscular atrophy (SMA) is an autosomal recessive neuromuscular disease and the most common cause of infant death worldwide, with an incidence of 1:10,000 live births and carrier frequency of 1:50.¹ This disease, caused by degeneration of spinal and bulbar motor neurons, is characterized by progressive muscle weakness and atrophy, scoliosis and feeding and respiratory problems.

Unravelling the complexity of SMA was a collaboration between insightful clinicians and laboratory scientists. The clinical spectrum includes observations reaching back over 100 years.² The first description, attributed to Werdnig in 1891, included two brothers aged 10 months.³ Hoffmann's 1893 review added seven cases of his own.⁴ A coincident description by Thomson and Bruce, also in 1893, brought a disease of intermediate severity with prominent scoliosis into the clinical spectrum.⁵ There was then a long hiatus until 1956 when Kugelberg and Welander described the third major clinical variant of SMA, previously thought to be a form of muscular dystrophy.⁶ Four decades later, in 1991, the International SMA Consortium on Childhood SMA classified patients into the three clinical groups that we recognize today (*Table 1*).^{7,8}

The acute form of type 1 SMA (Werdnig–Hoffmann disease) is characterized by severe generalized muscle weakness and hypotonia at birth or within the first 6 months, and is usually followed by death within 2 years. Children with type 2 SMA (Dubowitz disease) can sit, although they cannot stand or walk unaided, and survive beyond 2 years. In type 3 SMA (Kugelberg–Welander disease), patients have proximal muscle weakness, starting after the age of 18 months. In practical terms, clinical severity shows a continuous spectrum from mild to very severe SMA.⁹

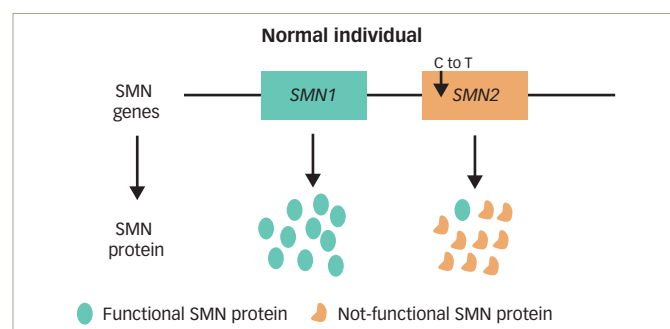
The unknown underlying biochemical defect made identifying the gene for SMA more challenging. Shortly after the clinical phenotypes were identified, Melki et al. mapped the chromosomal linkage of the three forms of SMA to chromosome 5 (5q11.2–13.3).^{10,11} Further characterization of the SMA locus revealed highly homologous duplications of the *SMN1* telomeric and *SMN2* centromeric regions. Both genes are transcribed. Five nucleotides distinguish the paralogous *SMN* genes and account for the alternative splicing with loss of exon 7 specific to *SMN2* transcripts (*Figure 1*).^{12,13} Full-length transcripts are almost exclusively produced by *SMN1*. Lefebvre et al. discovered that *SMN1* was lacking in 98% of patients with SMA.¹² *SMN2* is unable to compensate for the *SMN1* loss. Therefore, the development of the SMA phenotype is caused by two events: inherited (or *de novo*) *SMN1* gene mutations; and a constitutive defect of the *SMN2* gene leading to less full-length *SMN* protein being produced.¹²

Unravelling the clinical phenotypes and genetics of SMA represent the first steps in optimizing newborn screening and the treatment of this complex disease.

Table 1: Criteria for the classification of childhood spinal muscular atrophy, established in 1991⁷

Type*	Age at onset, months	Motor milestones	Age at death
I	<6	Never sit independently	<2 years
II	<18	Never walk independently	>2 years
III	>18	Stand and walk independently	Adult

*Since this classification, there have been two additions to chromosome 5-linked SMA. SMA type 0 is apparent in the pre-natal period as reduced foetal movement, and patients have only one copy of the survival motor neuron 2 (SMN2) gene. SMA type 4 is adult-onset disease usually manifesting in a patient's fourth decade of life. SMA = spinal muscular atrophy; SMN = survival motor neuron.

Figure 1: Schematic of survival motor neuron (SMN) genes on chromosome 5q13^{12,13}

There are two copies of the SMN gene, SMN1 and SMN2. A single nucleotide change in exon 7 of SMN2 results in exclusion of exon 7 (cytosine-to-thymine [C to T] mutation). Only 10% of normal SMN protein (grey) is produced by SMN2. Under normal conditions, almost all SMN protein is produced from SMN1.^{12,13} The SMN protein expressed from SMN2 is minimally functional (orange). SMN = survival motor neuron.

Efforts to treat spinal muscular atrophy based on antisense oligonucleotide technology

Following gene discovery, the first molecular efforts to treat SMA focused on correcting the SMN2 gene to compensate for the non-functional SMN protein. A single nucleotide mutation adversely affects splicing in exon 7 of SMN2. Blocking this mutation by antisense oligonucleotide therapy restores SMN2 during pre-mRNA splicing.¹⁴ Proof-of-concept studies showed efficacy in a transgenic mouse (D7) expressing human SMN2. The SMNΔ7 neonatal mouse model is also genetically *smn* null and has a lifespan of ~15 days.^{15,16} This enabled clinical trials of the antisense oligonucleotide nusinersen.^{17,18}

In the randomized, double-blind, sham-controlled, phase III ENDEAR trial, nusinersen was administered by intrathecal injection.¹⁷ Four loading doses were given and the first three doses administered at 14-day intervals.¹⁷ The fourth dose was administered 30 days after the third dose. Continued dosing was maintained every 4 months for the duration of the 13-month trial. The sham procedure consisted of a small needle prick to the skin over the lumbar spine. A total of 122 symptomatic infants were randomized; 81 were assigned to the nusinersen group, and 41 to the control group. All had weakness onset by 6 months.

The trial had two primary efficacy endpoints: a motor milestone response, which was defined according to results on the Hammersmith Infant Neurological Examination (HINE) (Section 2, HINE-2; scoring patient according to motor skills on eight items [Table 2]^{18,19} and event-free survival.) This was defined as the time to death or the use of permanent assisted ventilation (tracheostomy or ventilatory support for ≥16 h/day for >21 continuous days in the absence of an acute reversible event).¹⁸

At baseline, infants in the nusinersen group had earlier onset of symptoms and greater burden of disease compared with the control group. A pre-specified interim analysis at 2 years showed a significantly higher percentage of infants treated with nusinersen had a motor milestone response (41% versus 0% controls, $p < 0.001$). These results prompted early termination of the trial. In the final analysis, 51% of the infants in the nusinersen group and no infants in the control group had a motor milestone response. At final analysis, 39% of the infants in the nusinersen group and 68% in the control group had died or received permanent assisted ventilation. The median time to death or permanent assisted ventilation was 22.6 weeks in the control group and was not reached in the nusinersen group. Overall, the risk of death or the use of permanent assisted ventilation was 47% lower in the nusinersen group than in the control group ($p = 0.005$).¹⁷ Based on these results, on 23 December 2016, the US Food and Drug Administration (FDA) approved Spinraza® (nusinersen; Biogen, Durham, NC, USA) for all ages.²⁰

Risdiplam is an oral, small molecule, pre-mRNA splicing modifier that increases the production of the SMN protein, and in that way is related to the current discussion of nusinersen.^{19,21–24} Its efficacy results from its unique SMN2 pre-mRNA binding sites: a 5' splice site in intron 7 and exonic splicing enhancer in exon 7. This increases levels of full-length SMN mRNA and protein. Risdiplam has a significant advantage over nusinersen since it is the only oral medication approved for SMA,²⁵ though its oral dosing has potentially more advantages than just ease of administration. Upon absorption it will reach and be expressed in extraneuronal tissues where SMN protein is known to be deficient.²⁶ Such tissues include skeletal muscle, heart, bone, and autonomic and other nervous systems that may be contributing to disease state.²⁷ On 7 August 2020, the FDA approved Evrysdi® (risdiplam; Genentech, San Francisco, CA, USA) to treat patients 2 months and older with SMA. There is limited space in this review to fully discuss this compound but efficacy and safety have been established in a series of clinical trials.^{19,21–24} The application of risdiplam in the clinic will be further commented on in the *Conclusions*.

Gene replacement therapy

The path to gene replacement therapy for SMA was developed at the Nationwide Children's Hospital in Columbus, OH, USA. Two key factors enabled clinical trials in this area: the SMNΔ7 mouse model was available, and adeno-associated virus serotype 9 (AAV9) was shown to target neonatal neurons following intravascular delivery.²⁸ Preclinical studies demonstrated that self-complementary (sc)AAV9 infused on post-natal Day 1 rescued SMNΔ7 pups. Survival was increased from 15.0 days to 28.5 days with vector dosing of 6.7×10^{13} vg/kg, and to more than 250.0 days with doses of 3.3×10^{14} vg/kg. Treatment on post-natal Day 5 showed only partial correction, and post-natal Day 10 had little effect.²⁹

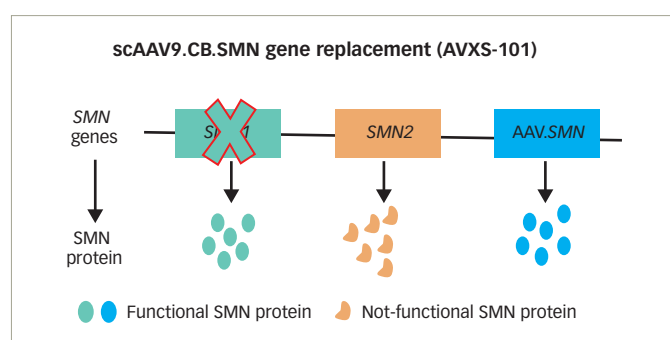
Pilot single-centre phase I/II clinical trial in spinal muscular atrophy type 1

Preclinical data defined the protocol for the first clinical trial, now named 'START', which started enrolling in 2014.³⁰ The treatment paradigm, as performed in the clinic, is shown in Figure 2. This was a dose-escalation trial in which patients were enrolled in two cohorts according to the dose administered. The primary outcome was safety, and the secondary outcome was time until death or the need for permanent ventilatory assistance. Exploratory outcomes included motor milestone achievements (particularly, sitting unassisted) and scores from Children's Hospital of Philadelphia Infant Test of Neuromuscular Disorders (CHOP INTEND; scores patients according to motor skills on 16 items; Table 3).^{17–19,21,22,24,30–32}

Table 2: The Hammersmith Infant Neurological Examination scoring system – Section 2: Motor milestones^{18,19}

Head control	Unable to maintain upright	Wobbles	Maintained upright all the time		
Sitting	Cannot sit	With support at hips	Props	Stable sit	Pivots
Voluntary grasp	No grasp	Uses whole hand	Index finger and thumb	Pincer grasp	
Ability to kick	No kick	Kick horizontal	Upward	Touches leg	Touches toes
Rolling	No rolling	Roll to side	Prone to supine or supine to prone	Supine to prone and prone to supine	
Crawling	No head lift	On elbow	On outstretched hand	On abdomen	On hands and knees
Standing	Not supported	Weight supported	With support	Unaided	
Walking	No walking	Bouncing	Cruising	Walks independently	

The Hammersmith Infant Neurological Exam (HINE) was developed to longitudinally assess the acquisition of motor milestones in type I SMA. It contains three sections: Section 1, assessing neurologic signs; Section 2, assessing motor function; and Section 3, assessing state of behaviour. Sections 1 and 3 are scored on descriptive ratings. Section 2 (HINE-2) is composed of the above motor milestones: head control, sitting, voluntary grasp, ability to kick, rolling, crawling, standing and walking. Each milestone achieved is given a point value; higher values indicate improved function (max score 26). The HINE-2 was used to assess infants with SMA in prospective treatment trials.^{18,19}

Figure 2: The principle of gene replacement in spinal muscular atrophy

In patients with spinal muscular atrophy, SMN1, the primary source of SMN protein (grey), is dysfunctional because of gene mutation. The small amount of SMN protein produced by the SMN2 gene will not compensate for SMN1 loss. AAV can deliver the SMN gene to the nucleus and restore SMN protein levels. AAV = adeno-associated virus; SMN = survival motor neuron.

Fifteen patients were enrolled in the clinical trial planned for infants from newborns to 9 months of age. Of the 16 patients screened, one was excluded because of elevated AAV9 antibody titres >1:50. Cohort 1 (n=3, mean age 6.3 months) received a single intravenous dose of scAAV9.CB.SMN (AVXS-101), 6.7×10^{13} vg/kg body weight. The first patient developed elevated serum aminotransferases (alanine aminotransferase $31 \times$ upper limit of normal [ULN]; aspartate aminotransferase $14 \times$ ULN) meeting criteria for a serious adverse event (SAE; Figure 3).^{30,33} Consequently, the protocol was amended, with all subsequent patients treated with oral prednisolone 1 mg/kg/day for 30 days, starting 24 hours before gene delivery. The amendment also included a dose adjustment for Cohort 2 (n=12; mean age 3.4 months), which received a single-dose vector 2.0×10^{14} vg/kg – half that used in preclinical trials.³⁰ Such was the impact of these protocol amendment, that this starting dose, alongside prednisone as an immunosuppressant, has been used in most subsequent neuromuscular clinical gene therapy trials.³⁴

As of 7 August 2017, all the patients had reached age ≥ 20 months without needing permanent mechanical ventilation (historical controls showed >90% of patients were dependent on permanent mechanical ventilation or had died at this age).^{30,35} Eleven patients were able to speak, a milestone rarely achieved in infants with SMA type 1. All the patients had increased CHOP INTEND scores from baseline and maintained these changes during the trial (Figure 4).³⁰ Cohort 2 had a mean increase of

9.8 points at 1 month and 15.4 points at 3 months. Sitting was achieved in 11 of 12 patients in Cohort 2, and nine could sit for at least 30 seconds. Most gratifying was the clinically meaningful results of feeding (hand to mouth), talking and sitting for at least 30 seconds in 11 of 12 patients in Cohort 2.²⁷

Two findings were related to age at gene delivery. The oldest patient treated was 7.9 months, and achieved little or no measurable benefits, thus, defining the limits of efficacy. On the other hand, the two youngest patients, who had the highest CHOP INTEND scores at the time of enrollment, achieved the highest scores on CHOP INTEND (>60 points) at the conclusion of the trial, and were able to crawl, stand and walk (Figure 4).³⁰ Both of these patients' families were alerted to SMA prior to delivery because of family history.³⁰ These outcomes strongly encourage early treatment and newborn screening for SMA.

The START trial was groundbreaking and defied all predictions about safety and efficacy issues for gene therapy. All 15 patients surpassed the previously reported median age of survival,³⁵ and motor milestones and clinically meaningful outcomes never before seen were achieved in this clinical trial.³⁰ As a result, the FDA approved onasemnogene abeparvovec gene therapy for SMA on 24 May 2019, under the commercial name Zolgensma® (Novartis, Durham, NC, USA). Multiple important follow-up clinical trials have been sponsored by Novartis, adding to the use of onasemnogene abeparvovec.^{31,36}

Phase III trial confirming efficacy and safety in spinal muscular atrophy type 1

A confirmatory, 12 centre, open-label, phase III trial (STR1VE) followed.³⁶ Eligibility criteria included patients aged ≤ 6 months with one or two copies of SMN2. Like START, this was a single intravenous dose of onasemnogene abeparvovec at a dose equivalent titre (1.1×10^{14} vg/kg). Co-primary efficacy endpoints were independent sitting for ≥ 30 seconds per Bayley-III at 18 months and survival (absence of death or permanent ventilation) at age 14 months. Historical controls from the Pediatric Neuromuscular Clinical Research dataset (PNCr) represented the comparator group.³⁶

Twenty-five patients were screened for AAV9 antibodies, and three were excluded with higher titres. The mean age at gene transfer was 3.7 months (12 females and 10 males), and none required feeding or ventilatory support. Thirteen (59.1%) patients achieved independent sitting ≥ 30 seconds at 18 months versus none in the control group ($p < 0.0001$). At age 18 months, 18 patients did not use ventilatory support versus no PNCr controls ($p < 0.0001$). Of the seven patients

Table 3: The Children's Hospital of Philadelphia Infant Test of Neuromuscular Disorders^{17-19,21,22,24,30-32}

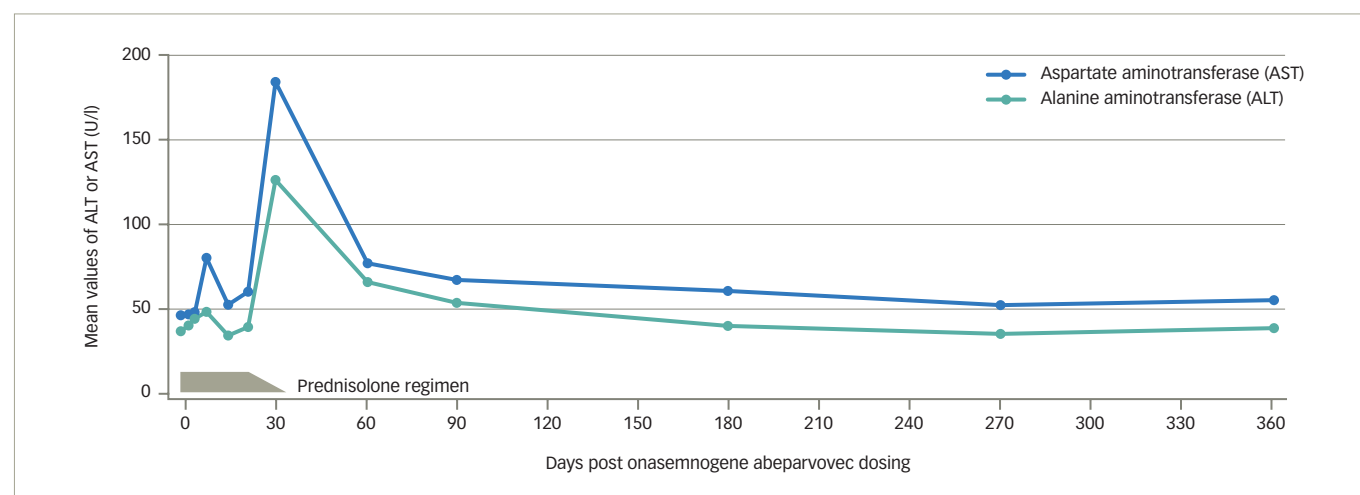
CHOP INTEND Item	Position	Graded response	Score
1 Spontaneous movement (upper extremity)	Supine	Antigravity shoulder movement (achieves elbow off surface)	4
		Antigravity elbow movement (achieves hand and forearm off surface)	3
		Wrist movement	2
		Finger movement	1
		No movement of limbs	0
2 Spontaneous movement (lower extremity)	Supine	Antigravity hip movement (achieves feet and knees off surface)	4
		Antigravity hip adduction/internal rotation (knees off surface)	3
		Active gravity eliminated knee movement	2
		Ankle movement	1
		No movement of limbs	0
3 Hand grip	Supine	Maintains hand grip with shoulder off bed	4
		Maintains grip with elbow off surface (shoulders on surface)	3
		Maintains grip with forearm off surface (elbow supported on surface)	2
		Maintains grip only with no traction	1
		No attempt to maintain grasp	0
4 Head in midline with visual stimulation	Supine head midline	Rotates from maximum rotation to midline	4
		Turns head part way back to midline	3
		Maintains midline for 5 or more seconds	2
		Maintains midline, >5 seconds	1
		Head falls to side, no attempts to regain midline	0
5 Hip adductors	Supine, no diaper	Keeps knee off surface of bed >5 seconds or lifts foot off surface	4
		Keeps knees off surface of bed 1-5 seconds	2
		No attempt to maintain knees off surface	0
6 Rolling (elicited from legs)	Supine (arms at side). Keep side tested up roll from the side tested	When traction is applied at the end of the manoeuvre, rolls to prone with lateral head righting	4
		Rolls through side-lying into prone without lateral head righting, clears weight-bearing arm to complete roll	3
		Pelvis, trunk and arm lift from support surface, head turns and rolls onto side, arm comes through to front of body	2
		Pelvis and trunk lift from support surface and head turns to side. Arm remains behind trunk	1
		Pelvis lifted passively off support surface	0
7 Rolling (elicited from arms)	Supine (arms at side). Keep side tested up roll from the side tested	Rolls to prone with lateral head righting	4
		Rolls into prone without lateral head righting; must clear weight-bearing arm completely to finish roll	3
		Rolls onto side, leg comes through and adducts, bringing the pelvis vertical	2
		Head turns to side and shoulder and trunk lift from surface	1
		Head turns to side; body remains limp or shoulder lifts passively	0
8 Shoulder and elbow flexion and horizontal abduction	Side-lying with upper arm at 30° of shoulder extension and elbow flexion and supported on body (restrain lower arm if needed)	Clears hand from surface with antigravity arm movement	4
		Able to flex shoulder to 45°, without antigravity arm movement	3
		Flexes elbow after arm comes off body	2
		Able to get arm off body	1
		No attempt	0
9 Shoulder flexion and elbow flexion	Sitting in lap with head and trunk support (20° recline)	Abducts or flexes shoulder to 60°	4
		Abducts or flexes shoulder to 30°	3
		Any shoulder flexion or abduction	2
		Flexes elbow only	1
		No attempt to lift arm	0

Table 3: Continued

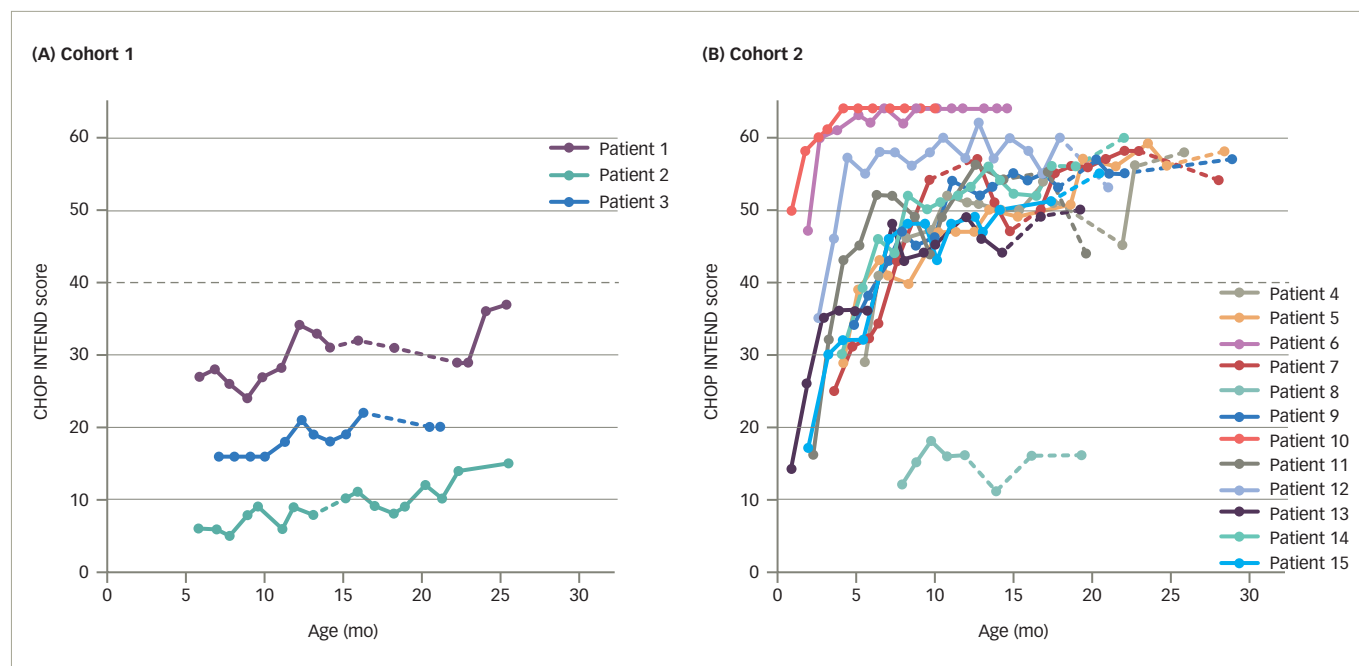
CHOP INTEND Item	Position	Graded response	Score
10 Spontaneous movement (lower extremity)	Sitting in lap or over edge of mat with head and trunk support (20° recline) thigh horizontal to ground	Extends knee to >45°	4
		Extends knee 15–45°	2
		Any visible knee extension	1
		No visible knee extension	0
11 Hip flexion and foot dorsiflexion	Hold infant against your body with legs free, facing outward. Support at the abdomen with child’s head resting between your arm and thorax	Hip flexion or knee flexion >30°	4
		Any hip flexion or knee flexion	3
		Ankle dorsiflexion only	2
		No active hip, knee or ankle motion	0
12 Head control	Sitting with support at the shoulders and trunk erect	Attains head upright from flexion and turns head side to side	4
		Maintains head upright for >15 s (for bobbing head control score a 2)	3
		Maintains head in midline for >5 s with the head tipped in up to 30° of forward flexion or extension	2
		Actively lifts or rotates head twice from flexion within 15 s (do not credit if movement is in time with breathing)	1
		No response, head hangs	0
13 Elbow flexion (score with item 14)	Supine	Flexes elbow	4
		Visible biceps contraction without elbow flexion	2
		No visible contraction	0
14 Neck flexion (score with item 13)	Supine	Lifts head off bed	4
		Visible muscle contraction of sternocleidomastoid	2
		No muscle contraction	0
15 Head/neck extension (Landau)	Ventral suspension: Prone, held in one hand upper abdomen	Extends head to horizontal plane or above	4
		Extends head partially, but not to horizontal	2
		No head extension	0
16 Spinal incurvation (Galant)	Ventral suspension: Prone, held in one hand upper abdomen	Twists pelvis towards stimulus off axis	4
		Visible paraspinal muscle contraction	2
		No response	0

The Children’s Hospital of Philadelphia Infant Test of Neuromuscular Disorders (CHOP INTEND) objectively assesses motor milestones with a scoring system across 16 outcome measures. It is validated as a test of infant development and used in multiple spinal muscular atrophy clinical trials.^{17–19,21,22,24,30–32} Each of the 16 outcomes is scored from 0 to 4 (with 0 being inability to perform the movement and 4 being able to perform the task; 0–64 points).

Figure 3: Effect of onasemnogene abeparvovec on liver enzymes in the START trial^{30,33}



Mean values for liver enzymes (AST and ALT) that increased 3–4 weeks post-gene delivery in the START study. Prednisolone brought this back to baseline. A protocol amendment led to all subsequent patients receiving oral prednisolone 1 mg/kg/day for 30 days, starting 24 hours before gene delivery.³⁰ ALT = alanine aminotransferase; AST = aspartate aminotransferase. Reproduced with permission from Al-Zaidy and Mendell. 2019.³³

Figure 4: Children's Hospital of Philadelphia Infant Test of Neuromuscular Disorders scores of patients in the START trial³⁰

Children's Hospital of Philadelphia Infant Test of Neuromuscular Disorders (CHOP INTEND) scores (maximum 64 points) for: (A) Cohort 1 ($n=3$) treated with low-dose vector (6.7×10^{13} vg/kg); and (B) Cohort 2 ($n=12$) treated with high-dose vector (2.0×10^{14} vg/kg). Patients with high CHOP INTEND scores at baseline and who receive early treatment achieve the most favourable outcome. Patients with late dosing and a low CHOP INTEND score at baseline had poor outcomes.³⁰ The black dashed lines indicate the maximum score reached by historical SMA type 1 controls.

CHOP INTEND = Children's Hospital of Philadelphia Infant Test of Neuromuscular Disorders
Reproduced with permission from Mendell, et al. (2017).³⁰

using non-invasive ventilation, five had documented previous use of Trilogy 100 and two had used other types of non-invasive ventilation. In addition, a clinically meaningful measure of ability to thrive at 18 months included three assessments: (1) ability to tolerate thin liquids by clinical swallowing assessment (55%); (2) able to feed exclusively by mouth (86%); and (3) weight maintained and consistent with age (14%).³⁶

Regarding CHOP INTEND scores, there was a mean increase from baseline of 6.9 points at 1 month post-dosing, 11.7 points at 3 months and 14.6 points at 6 months.³⁶ Twenty-one (95%) patients achieved a CHOP INTEND score of ≥ 40.0 points, 14 (64%) achieved ≥ 50.0 points and five (23%) achieved ≥ 60.0 points. Historically, children with SMA type 1 almost never achieve CHOP INTEND scores >40 points.³⁵

The safety profile demonstrated in STR1VE was again remarkable and convincing for continued use of onasemnogene abeparvovec. All adverse events were documented, but only three (14%) SAEs were related to gene delivery. Two of these SAEs were elevated liver enzymes, a finding consistent with other AAV clinical trials. The third SAE was hydrocephalus, occurring for unknown reasons. This trial validated the findings of the START trial and opened avenues for gene therapy for many other childhood diseases.

Gene therapy for spinal muscular atrophy type 2

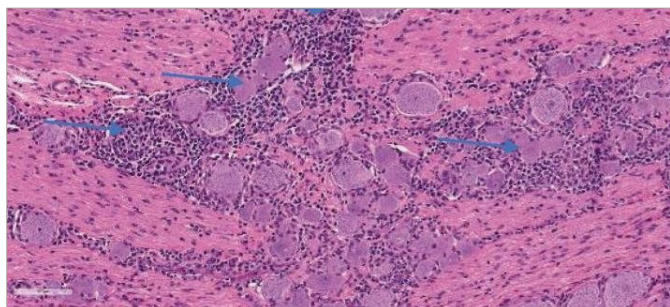
Considering the success of onasemnogene abeparvovec in treating infants with SMA type 1, on-going studies are addressing its potential in treating patients with sitting, non-ambulatory SMA type 2, and SMA type 3. Treatment at an older age would require higher intravenous dosing of onasemnogene abeparvovec due to body weight, accompanied by a higher risk of side effects. To circumvent these obstacles, gene transfer directly targeting the bulbar and spinal cord motor neurons was planned

by intrathecal delivery. Preclinical studies showed that intrathecal delivery to mice and non-human primates was safe and effective at a viral vector tenfold lower than with IV dosing.^{37,38}

The phase I, open-label, ascending-dose STRONG study started in December 2017.³⁹ Patients with SMA type 2, aged between 6 and <12 months, with three copies of *SMN2*, who were able to sit unassisted for 10 seconds but unable to walk or stand were included. Those with severe scoliosis ($\geq 50^\circ$ curve) were excluded. There were three onasemnogene abeparvovec dosing cohorts: Cohort 1, receiving 6.0×10^{13} vg; Cohort 2, receiving 1.2×10^{14} vg; and Cohort 3, receiving 2.4×10^{14} vg. Each group was stratified by age, either 6 to <24 months and 24 to <60 months. The primary endpoints were safety/tolerability, independent standing for ≥ 3 seconds in patients aged 6 to <24 months or change in Hammersmith Functional Motor Scale–Expanded score in patients aged 24 to <60 months. Outcomes were compared with those of PNCR historical controls. Patients received prophylactic prednisolone (1 mg/kg/day) 24 hours prior to intrathecal delivery, maintained for approximately 30 days with a taper depending on clinical toxicity. Onasemnogene abeparvovec was delivered as a single intrathecal injection, with the patient sedated and in the Trendelenburg position at 30° to enhance distribution to the spinal cord and brain.

In October 2019, as enrolment was complete for Cohort 1 ($n=3$) and Cohort 2 ($n=25$) but incomplete for Cohort 3 ($n=4$), the FDA imposed a partial clinical hold on the study resulting from a Novartis report to the FDA of unexpected findings in the dorsal root ganglion (DRG) in onasemnogene abeparvovec-treated non-human primates.⁴⁰ This preclinical study showed mononuclear cell inflammation, sometimes accompanied by neuronal cell body degeneration or loss (Figure 5), though the non-human primates were asymptomatic.³⁸ Novartis then did further research to assess clinical

Figure 5: Dorsal root ganglion from a non-human primate after treatment with intrathecal onasemnogene abeparvovec³¹



Dorsal root ganglion from a non-human primate dosed at 3×10^{13} vg/animal by intrathecal lumbar puncture shows moderate neuron degeneration (arrows) and mononuclear cell inflammation 6 weeks post-gene delivery. Hematoxylin and eosin stain; magnification 200x.

Reproduced with permission from Tukov FF, et al. 2022.³¹

impact. Intrathecal delivery of vector in additional studies showed DRG and trigeminal ganglion inflammation with scattered neuronal degeneration. The findings at 52 weeks post-dosing were non-progressive and were of minimal severity compared to interim autopsy data performed at earlier time points.³⁸ In August 2021, the FDA determined that the STRONG study could proceed with intrathecal delivery.⁴¹

Despite release from clinical hold, the sponsor elected not to enrol more patients. Safety issues appeared minimal following intrathecal delivery and the SAEs related only to transaminase elevations without increase in bilirubin. No signs of DRG toxicity were encountered.³⁹

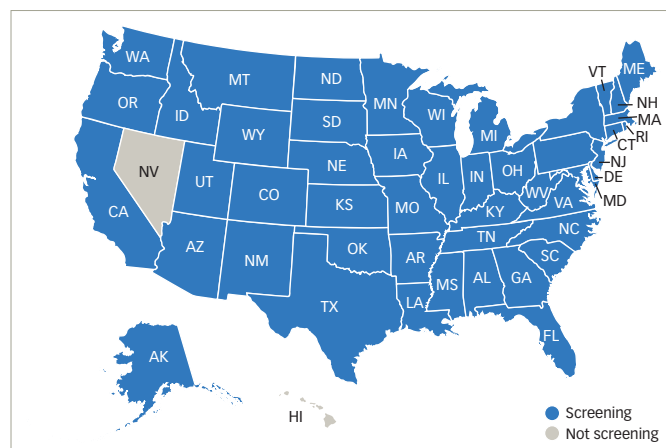
Further studies are needed to validate the efficacy of intrathecal delivery in SMA type 2. To address this, Novartis is sponsoring STEER, a randomized, sham-controlled, double-blind phase III study (Efficacy and safety of intrathecal OAV101 [AVXS-101] in pediatric patients with type 2 spinal muscular atrophy (SMA); ClinicalTrials.gov identifier: NCT05089656).⁴² In the STEER study, treatment-naïve patients with SMA type 2, aged 2–17 years, will be treated with intrathecal onasemnogene abeparvovec at 1.2×10^{14} vg, a safe dose used in the STRONG trial.

Impact of gene therapy for newborn infants with spinal muscular atrophy

SMA was added to the Department of Health and Human Services' Recommended Uniform Screening Panel in February 2018, a little more than a year after nusinersen was approved for SMA treatment and three months after the START study in SMA type 1 gene therapy was published.^{30,43} Currently 48 states include SMA in their newborn screening panel (Figure 6).⁴⁴

The value of onasemnogene abeparvovec is unequivocal for treating newborns, evidenced by a phase III, multicentre, single-arm trial (SPR1NT).^{31,32} SPR1NT included infants with pre-symptomatic SMA, with biallelic *SMN1* deletions, and two or three copies of *SMN2*. Other newborns were excluded because of signs of SMA, reduced compound muscle action potentials and elevated AAV9 antibody titres. Infants with two copies were treated at median age of 21 days (range 8–34 days), and those with three copies were treated at median age of 32 days (range 9–43); the first was enrolled in April 2018.^{31,32}

Figure 6: US states currently screening or not screening for spinal muscular atrophy⁴⁴



In the USA, 48 states currently screen for SMA, and 98% of newborns are screened and have an opportunity for gene therapy.

Map reproduced with permission of CureSMA.⁴⁴

For the cohort of 14 infants with two copies of *SMN2*, the efficacy and safety of onasemnogene abeparvovec was demonstrated, and outcomes were remarkably consistent given that all 14 patients achieved the primary endpoint of independent sitting for at least 30 seconds ($p < 0.0001$). All 14 (100%) achieved motor milestones assessed by Bayley-III, and 11 of 14 stood alone and walked independently. All children in this cohort of the SPR1NT trial achieved a CHOP INTEND score of at least 58 by 18 months of age. None required any form of respiratory or feeding support.³¹

Results from the second cohort of patients, those with three copies of *SMN2*, are important in translating the findings into clinical practice because most infants with this molecular profile will develop clinical manifestations consistent with SMA type 2 or 3. Three copies of *SMN2* predicts onset of SMA type 2 at age 7–18 months in 54% of patients, and type 3 phenotype in 31%. Patients with SMA type 2 can sit independently, some can stand and none can walk.³²

For the cohort of infants with three copies of *SMN2*, 15 infants (13 identified by newborn screening) were analysed. All 15 patients were able to stand independently for at least 3 seconds at a median age of 377 days, 14 patients walked independently by 24 months of age, and all patients had Bayley Scales of Infant and Toddler Development motor scores comparable to neurologically 'normal' infants. All survived, without respiratory or nutritional support by trial end.³²

The safety profile of early intravenous treatment was exceptional in both cohorts of patients in SPR1NT. In patients with two copies, mild hepatotoxicity was observed in only three subjects. Troponin-I elevation was rarely encountered and there was no compromised cardiac function by echocardiogram. Platelet counts remained $>75,000$ for all infants throughout the study.^{31,32}

Commercial gene transfer treatment of 21 infants from Ohio confirmed the lessons learned from SPR1NT.⁴⁵ Infants who were recognized by newborn screening and were treated early fared the best. Children older than 6 months were more likely to have asymptomatic transaminase elevation and were treated with longer immunosuppression.⁴⁵ The safety information for Zolgensma[®] is summarized in Table 4, including reports on recent deaths.^{46,47}

Table 4: Important Safety Information as part of the Prescribing Information for Zolgensma® (onasemnogene abeparvovec)^{46,47}

Acute serious liver injury and acute liver failure
<ul style="list-style-type: none"> Acute serious liver injury, acute liver failure and elevated aminotransferases can occur Patients with pre-existing liver impairment may be at higher risk Prior to gene delivery, check clinical and laboratory signs including aminotransferases, total bilirubin and prothrombin time. Continue to monitor liver function for 3 months after infusion
Additional risks to consider
<ul style="list-style-type: none"> Thrombocytopenia: Typically occurs in the first 2 weeks post-infusion; platelets should be monitored before and after infusion and on a regular basis for 3 months Thrombotic microangiopathy: Greatest risk approximately 1 week post-infusion. Obtain platelet counts, creatinine and complete blood counts before and after infusion. Specialists may be needed for haematologic or renal complications when present Elevated troponin-I: Cardiac troponin may be observed post-infusion. Monitor troponin-I before infusion and regularly for 3 months
Adverse reactions
The most common adverse reactions (≥5%) in clinical studies were elevated aminotransferases and vomiting
Deaths related to Zolgensma®
Two deaths caused by acute liver failure were reported on in August 2022. ⁴⁷ The first fatalities directly related to Zolgensma® occurred 5–6 weeks post-infusion. Deaths occurred following tapering of steroids that had been given to suppress immune reaction following gene delivery

Conclusion

The results of molecular-based treatment for SMA type 1, when used prior to onset of symptoms, are overwhelmingly successful for a disease predicting death by age 2 years. Treatment outcomes in infants with SMA type 1 vary depending on age and severity of disease at gene transfer. The FDA has approved intravenous systemic gene delivery for patients up to 2 years of age, and for pre-symptomatic infants, we have observed sustained benefit for nearly 5 years.⁴⁸ Results of treatment of SMA type 2 require further evaluation.

For patients still showing manifestations post-gene delivery, it is acceptable to consider combination treatment with other approved therapies for SMA (nusinersen or risdiplam). There are no clinical trials directly addressing this question, but favourable results have been reported with gene transfer and nusinersen without an increase in adverse events.^{49,50} Thus, patients

receiving gene therapy beyond 6 months of age with residual signs of disease following treatment might benefit by receiving nusinersen. Similar studies are not currently available with risdiplam.

In patients for whom first-line gene therapy is not possible (e.g. those with pre-existing AAV antibody), the choice between nusinersen and risdiplam may be difficult for physicians. There are no head-to-head trials comparing efficacy, but overall, the results support the use of risdiplam as an important alternative to nusinersen for the treatment of patients with SMA type 1.⁵¹

SMA gene therapy has been a leading example of safety and efficacy of gene therapy, especially for other infant genetic diseases. As a breakthrough treatment, it provides a path for gene delivery for older children and adults. □

- Hoyert DL, Xu J. Deaths: Preliminary data for 2011. *Natl Vital Stat Rep.* 2012;61:1–51.
- Dubowitz V. Ramblings in the history of spinal muscular atrophy. *Neuromuscul Disord.* 2009;19:69–73.
- Werdnig G. Zwei frühinfantile hereditäre fälle von progressiver muskeltrophie unter dem bilde der dystrophie, aber anf neuroischer grundlage. *Archiv für Psychiatrie und Nervenkrankheiten.* 1891;22:437–80.
- Hoffmann J. Ueber chronische spinale muskeltrophie im kindesalter, auf familiärer basis. *Deutsche Zeitschrift für Nervenheilkunde.* 1893;3:427–70.
- Thompson J, Bruce A. Progressive muscular atrophy in a child with a spinal lesion. *Edinb Hosp Rep.* 1893;1:372.
- Kugelberg E, Welander L. Heredofamilial juvenile muscular atrophy simulating muscular dystrophy. *AMA Arch Neurol Psychiatry.* 1956;75:500–9.
- Munsat T. Workshop report: International SMA collaboration. *Neuromuscul Disord.* 1991;1:181.
- Munsat T, Davies, KE. International SMA consortium meeting. *Neuromuscul Disord.* 1992;2:423–8.
- Arnold WD, Kassas D, Kissel JT. Spinal muscular atrophy: Diagnosis and management in a new therapeutic era. *Muscle Nerve.* 2015;51:157–67.
- Melki J, Sheth P, Abdelhak S, et al. Mapping of acute (type I) spinal muscular atrophy to chromosome 5q12-q14. The French spinal muscular atrophy investigators. *Lancet.* 1990;336:271–3.
- Melki J, Lefebvre S, Burglen L, et al. De novo and inherited deletions of the 5q13 region in spinal muscular atrophies. *Science.* 1994;264:1474–7.
- Lefebvre S, Burglen L, Rebollet S, et al. Identification and characterization of a spinal muscular atrophy-determining gene. *Cell.* 1995;80:155–65.
- Lorson CL, Hahnen E, Androphy EJ, et al. A single nucleotide in the SMN gene regulates splicing and is responsible for spinal muscular atrophy. *Proc Natl Acad Sci.* 1999;96:6307–11.
- Singh RN, Howell MD, Ottesen EW, et al. Diverse role of survival motor neuron protein. *Biochim Biophys Acta Gene Regul Mech.* 2017;1860:299–315.
- Hsieh-Li HM, Chang JG, Jong YJ, et al. A mouse model for spinal muscular atrophy. *Nat Genet.* 2000;24:66–70.
- Monani UR, Sendtner M, Coovert DD, et al. The human centromeric survival motor neuron gene (SMN2) rescues embryonic lethality in *Smn(-/-)* mice and results in a mouse with spinal muscular atrophy. *Hum Mol Genet.* 2000;9:333–9.
- Finkel RS, Mercuri E, Darras BT, et al. Nusinersen versus sham control in infantile-onset spinal muscular atrophy. *N Engl J Med.* 2017;377:1723–32.
- Finkel RS, Chiriboga CA, Vajsar J, et al. Treatment of infantile-onset spinal muscular atrophy with nusinersen: A phase 2, open-label, dose-escalation study. *Lancet.* 2016;388:3017–26.
- Baranello G, Darras BT, Day JW, et al. Risdiplam in type 1 spinal muscular atrophy. *N Engl J Med.* 2021;384:915–23.
- US Food and Drug Administration. FDA approves first drug for spinal muscular atrophy. 2016. Available at: www.fda.gov/news-events/press-announcements/fda-approves-first-drug-spinal-muscular-atrophy (accessed 17 October 2022).
- Darras BT, Masson R, Mazurkiewicz-Beldzinska M, et al. Risdiplam-treated infants with type 1 spinal muscular atrophy versus historical controls. *N Engl J Med.* 2021;385:427–35.
- Mercuri E, Deconinck N, Mazzone ES, et al. Safety and efficacy of once-daily risdiplam in type 2 and non-ambulant type 3 spinal muscular atrophy (SUNFISH part 2): A phase 3, double-blind, randomised, placebo-controlled trial. *Lancet Neurol.* 2022;21:42–52.
- Sergott RC, Amorelli GM, Baranello G, et al. Risdiplam treatment has not led to retinal toxicity in patients with spinal muscular atrophy. *Ann Clin Transl Neurol.* 2021;8:54–65.
- Mercuri E, Baranello G, Boespflug-Tanguy O, et al. Risdiplam in types 2 and 3 spinal muscular atrophy: A randomised, placebo-controlled, dose-finding trial followed by 24 months of treatment. *Eur J Neurol.* 2022. doi: 10.1111/ene.15499.
- US Food and Drug Administration. FDA approves oral treatment for spinal muscular atrophy. 2020. Available at: www.fda.gov/news-events/press-announcements/fda-approves-oral-treatment-spinal-muscular-atrophy (accessed 17 October 2022).
- Nash LA, Burns JK, Chardon JW, et al. Spinal muscular atrophy: More than a disease of motor neurons? *Curr Mol Med.* 2016;16:779–92.
- Simone C, Ramirez A, Bucchia M, et al. Is spinal muscular atrophy a disease of the motor neurons only: Pathogenesis and therapeutic implications? *Cell Mol Life Sci.* 2016;73:1003–20.
- Foust KD, Nurre E, Montgomery CL, et al. Intravascular AAV9 preferentially targets neonatal neurons and adult astrocytes. *Nat Biotechnol.* 2009;27:59–65.
- Foust KD, Wang X, McGovern VL, et al. Rescue of the spinal muscular atrophy phenotype in a mouse model by early postnatal delivery of SMN. *Nat Biotechnol.* 2010;28:271–4.
- Mendell JR, Al-Zaidy S, Shell R, et al. Single-dose gene-replacement therapy for spinal muscular atrophy. *N Engl J Med.* 2017;377:1713–22.
- Strauss KA, Farrar MA, Muntoni F, et al. Onasemnogene abeparvovec for presymptomatic infants with two copies of SMN2 at risk for spinal muscular atrophy type 1: The phase III SPR1NT trial. *Nat Med.* 2022;28:1381–9.
- Strauss KA, Farrar MA, Muntoni F, et al. Onasemnogene abeparvovec for presymptomatic infants with three copies of SMN2 at risk for spinal muscular atrophy: The phase III SPR1NT trial. *Nat Med.* 2022;28:1390–7.
- Al-Zaidy SA, Mendell JR. From clinical trials to clinical practice: Practical considerations for gene replacement therapy for SMA type 1. *Pediatr Neurol.* 2019;100:3–11.
- Mendell JR, Sahenk Z, Lehman K, et al. Assessment of systemic delivery of rAAVrh74.MHCK7.micro-dystrophin in children with duchenne muscular dystrophy. *JAMA Neurol.* 2020;77:1122–31.
- Finkel RS, McDermott MP, Kaufmann P, et al. Observational study of spinal muscular atrophy type 1 and implications for clinical trials. *Neurology.* 2014;83:810–7.
- Day JW, Finkel RS, Chiriboga CA, et al. Onasemnogene abeparvovec gene therapy for symptomatic infantile-onset spinal muscular atrophy in patients with two copies of SMN2 (STRIVE): An open-label, single-arm, multicentre, phase 3 trial. *Lancet Neurol.* 2021;20:284–93.
- Passini MA, Bu J, Richards AM, et al. Translational fidelity of intrathecal delivery of self-complementary AAV9-survival motor neuron 1 for spinal muscular atrophy. *Hum Gene Ther.* 2014;25:619–30.
- Tukov FF, Mansfield K, Milton M, et al. Single-dose intrathecal dorsal root ganglia toxicity of onasemnogene abeparvovec in cynomolgus monkeys. *Hum Gene Ther.* 2022;33:740–56.
- ClinicalTrials.gov. Study of Intrathecal Administration of Onasemnogene Abeparvovec-xioi for Spinal Muscular Atrophy (STRONG). ClinicalTrials.gov Identifier: NCT03381729. Available at: <https://clinicaltrials.gov/ct2/show/NCT03381729> (accessed 28 September 2022).
- Meyer K, Ferraiuolo L, Schmelzer L, et al. Improving single injection CSF delivery of AAV9-mediated gene therapy for SMA:

- A dose-response study in mice and nonhuman primates. *Mol Ther.* 2015;23:477–87.
41. Novartis. Novartis announces lift of partial clinical trial hold and plans to initiate a new, pivotal Phase 3 study of intrathecal OAV-101 in older patients with SMA. Available at: www.novartis.com/news/media-releases/novartis-announces-lift-partial-clinical-trial-hold-and-plans-initiate-new-pivotal-phase-3-study-intrathecal-oav-101-older-patients-sma (accessed 28 September 2022).
 42. ClinicalTrials.gov. Efficacy and Safety of Intrathecal OAV101 (AVXS-101) in Pediatric Patients With Type 2 Spinal Muscular Atrophy (SMA) (STEER). ClinicalTrials.gov Identifier: NCT05089656. Available at: <https://clinicaltrials.gov/ct2/show/NCT05089656> (accessed 28 September 2022).
 43. Cure SMA. Advisory committee on heritable disorders in newborns and children recommends nationwide newborn screening for spinal muscular atrophy. 2018. Available at: www.curesma.org/advisory-committee-on-heritable-disorders-in-newborns-and-children-recommends-nationwide-newborn-screening-for-spinal-muscular-atrophy/ (accessed 17 October 2022).
 44. Cure SMA. Newborn screening for SMA. 2022. Available at: www.curesma.org/newborn-screening-for-sma/ (accessed 28 September 2022).
 45. Waldrop MA, Karingada C, Storey MA, et al. Gene therapy for spinal muscular atrophy: Safety and early outcomes. *Pediatrics.* 2020;146:e20200729.
 46. US Food and Drug Administration. ZOLGENSMA® (onasemnogene abeparvec-xioi). Prescribing information. Available at: www.fda.gov/media/126109/download (accessed 17 October 2022).
 47. Liu A. 2 deaths after Novartis' Zolgensma put gene therapy's liver safety in the spotlight once again. 2022. Available at: www.fiercepharma.com/pharma/two-deaths-after-novartis-zolgensma-bring-gene-therapys-liver-safety-spotlight-again (accessed 17 October 2022).
 48. Mendell JR, Al-Zaidy S, Lehman K, et al. Five-year extension results of the phase 1 START trial of onasemnogene abeparvec in spinal muscular atrophy. *JAMA Neurol.* 2021;78:834–41.
 49. Harada Y, Rao VK, Arya K, et al. Combination molecular therapies for type 1 spinal muscular atrophy. *Muscle Nerve.* 2020;62:550–4.
 50. Lee BH, Collins E, Lewis L, et al. Combination therapy with nusinersen and AVXS-101 in SMA type 1. *Neurology.* 2019;93:640–1.
 51. Ribero VA, Daigi M, Marti Y, et al. How does risdiplam compare with other treatments for types 1–3 spinal muscular atrophy: A systematic literature review and indirect treatment comparison. *J Comp Eff Res.* 2022;11:347–70.